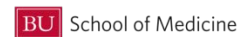


# Liquid Biopsy for Diagnosis and Treatment Monitoring in Esophageal Adenocarcinoma

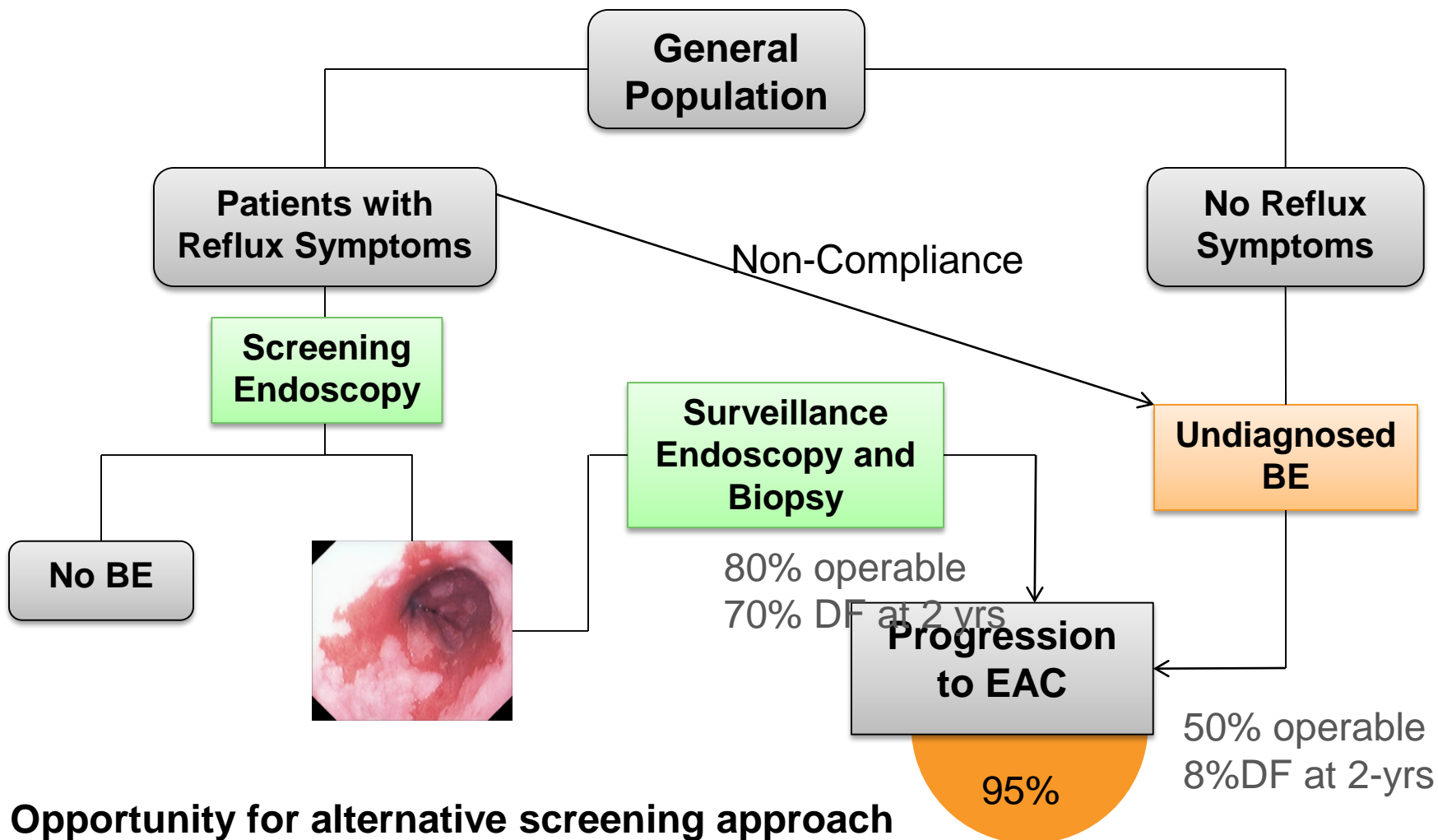
Tony E. Godfrey, PhD  
Associate Chair-Research  
Department of Surgery  
Boston University School of Medicine



**UPMC**  
University of Pittsburgh  
Medical Center



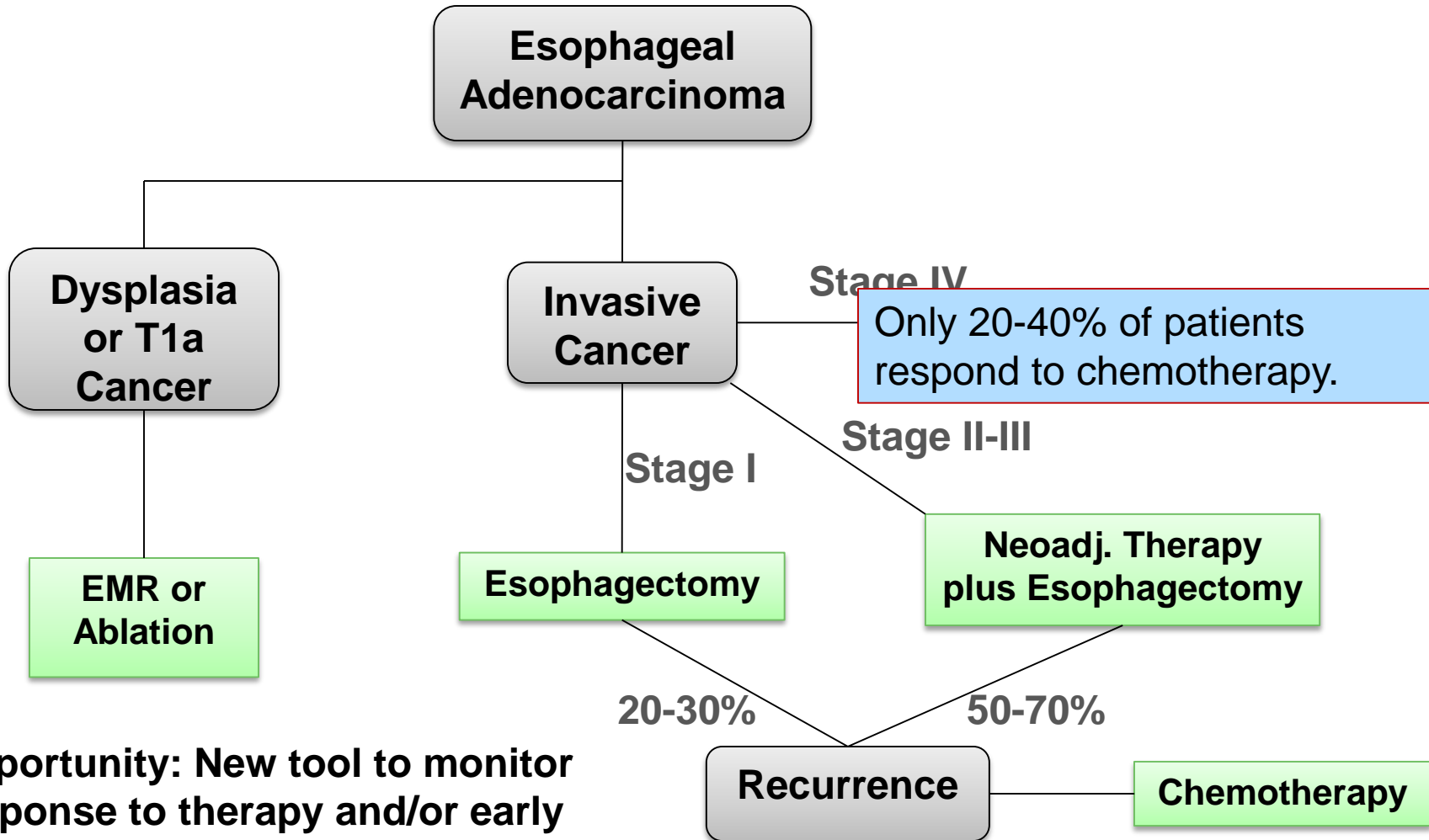
# Why ctDNA in Esophageal Adenocarcinoma?



Opportunity for alternative screening approach

Cancer

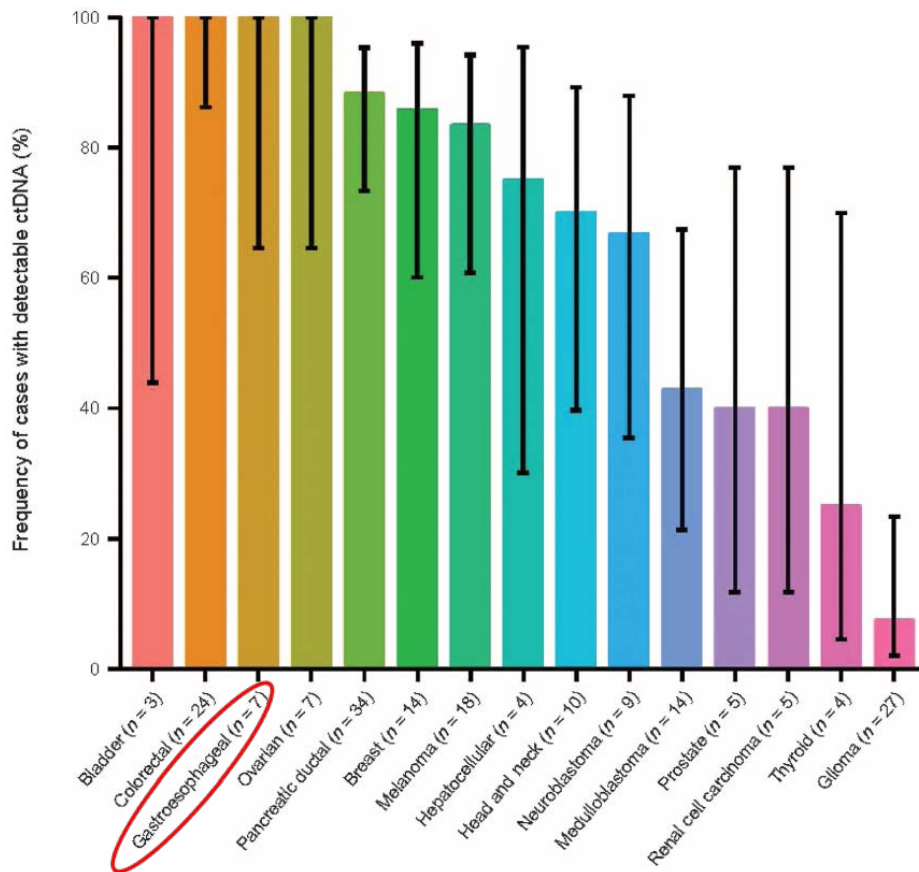
# Why ctDNA in Esophageal Adenocarcinoma?



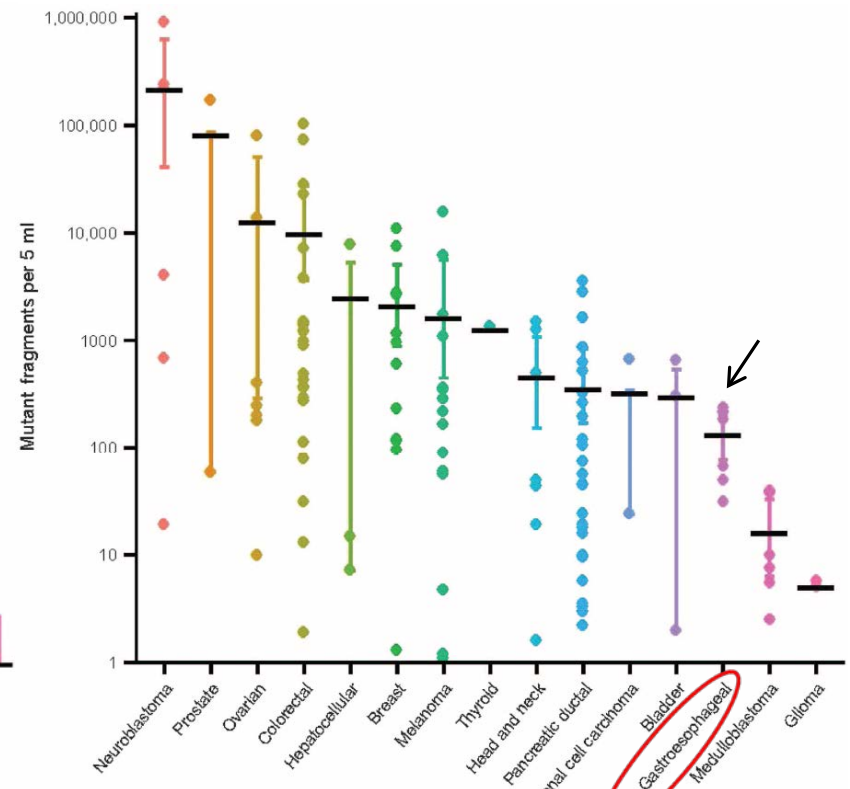
**Opportunity: New tool to monitor response to therapy and/or early detection of recurrence**

# Limited but Promising data on ctDNA in Gastroesophageal Cancer

ctDNA detected in 100% of metastatic cases (n=7)



ctDNA detected at ~10-100 copies/ml of plasma



Bettegowda *et al. Sci Transl Med* (2014)

# Circulating Tumor DNA in EAC

## ▪Open Questions:

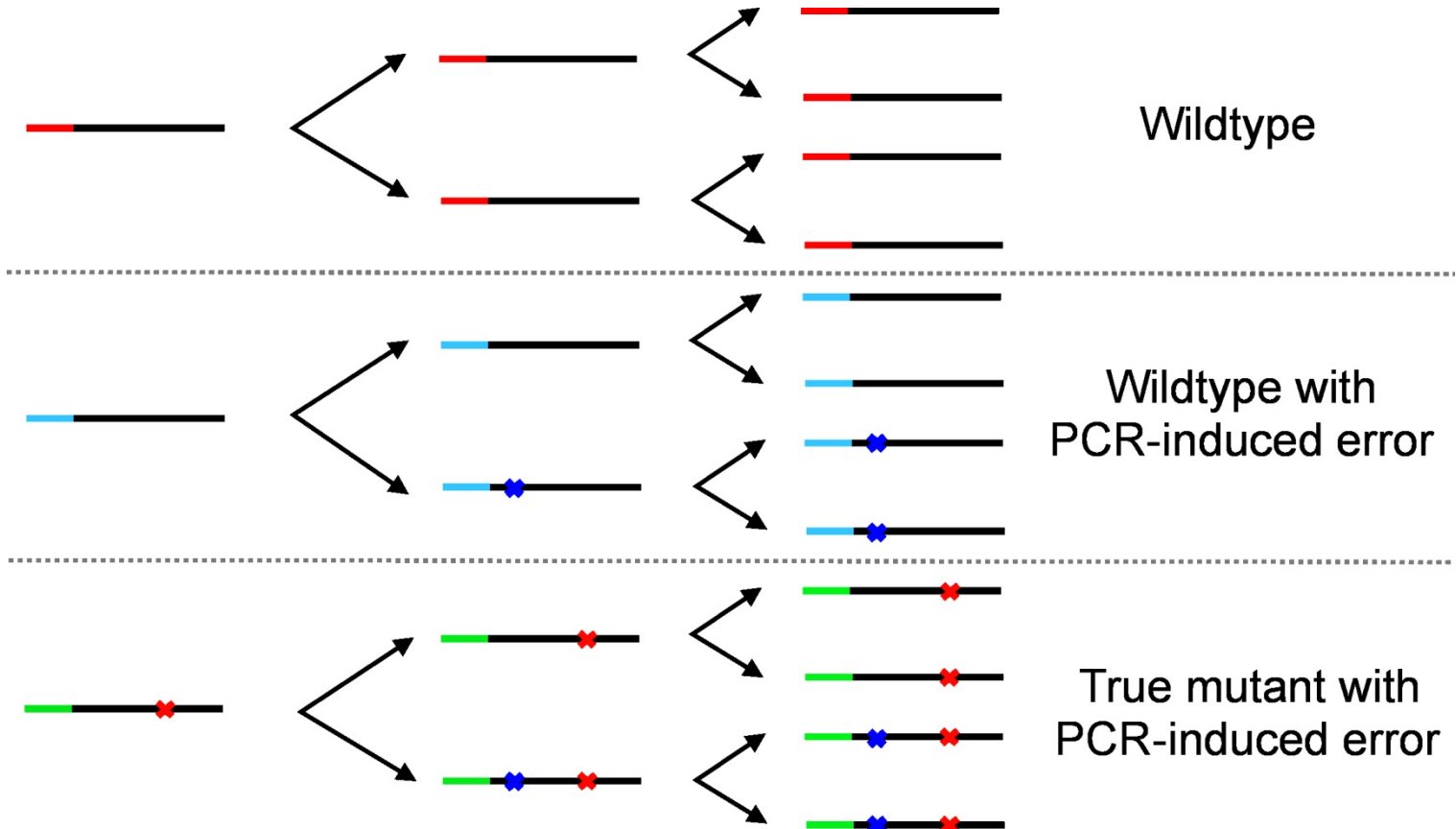
- How does detection rate and quantity change with tumor stage?
- Does ctDNA quantity change with response to therapy or disease progression/recurrence?

## ▪Challenges:

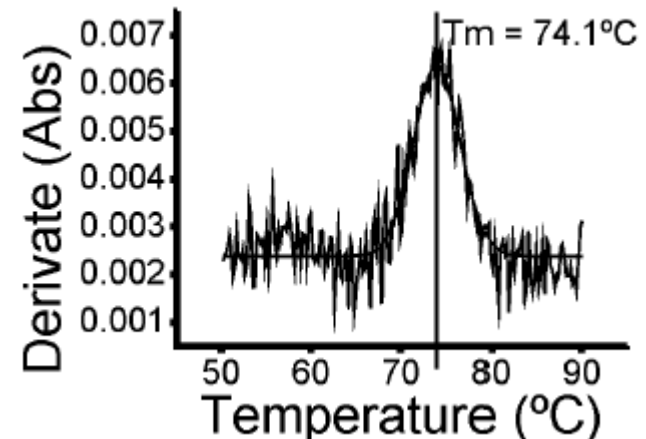
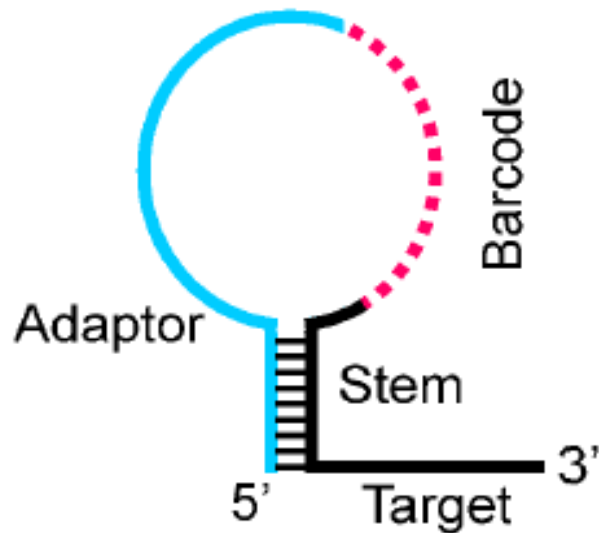
- Need to reliably detect mutations at ~0.05% level.
- Very little DNA in plasma/serum (10-50ng/mL = 1-10 copies of tumor DNA) and it is very fragmented.
- **Developed modified version of SafeSeq-S using introduction of molecular barcodes ( $N_{10-14}$ ) into NGS libraries using PCR.**

Kinde et al., Proc Natl Acad Sci U S A. 2011 Jun 7;108(23)

# Barcoding Enables Identification of True Mutations vs Polymerase errors



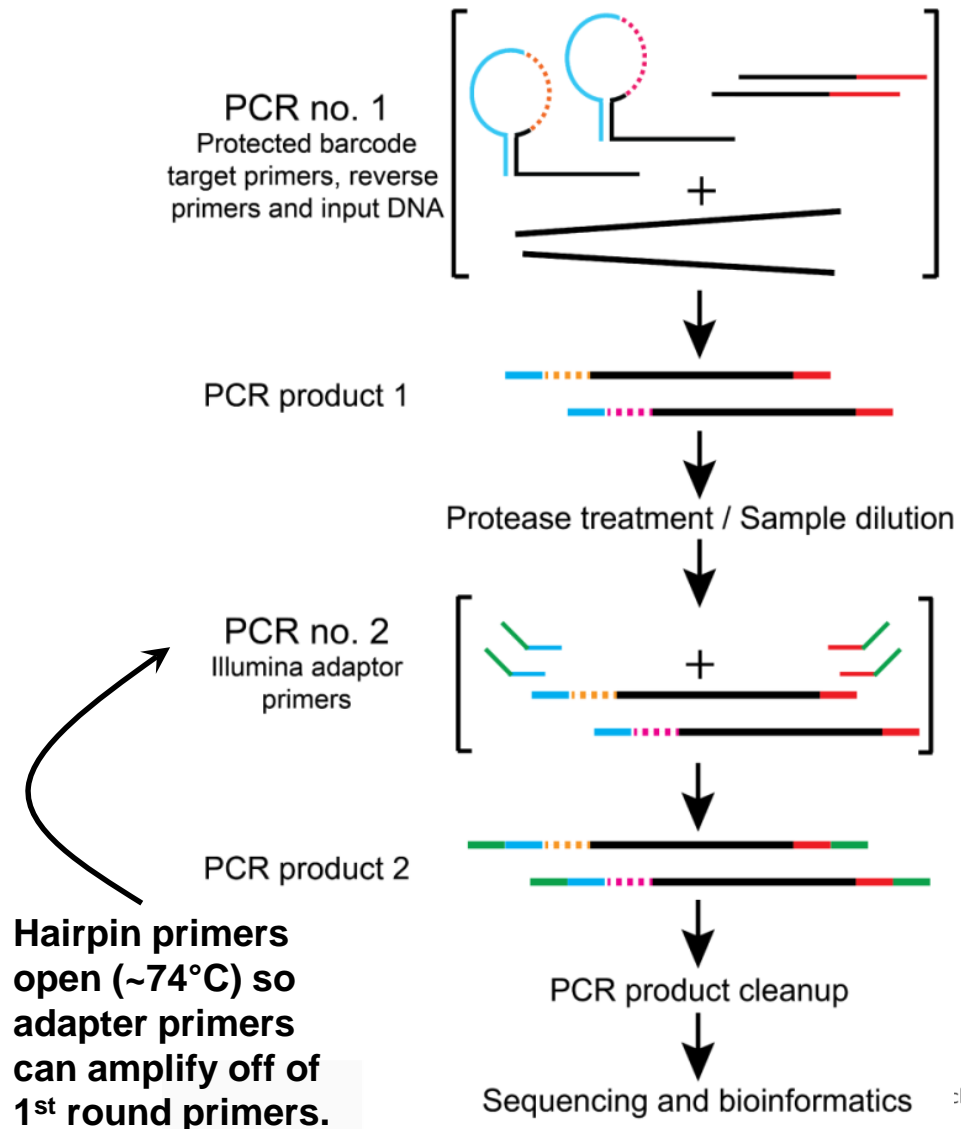
# Simple, Multiplexed, PCR-based barcoding of DNA for Sensitive mutation detection using Sequencing (SiMSen-Seq)



- Target primers designed with internal hairpin.
  - Protects random barcode sequence during PCR and increases specificity

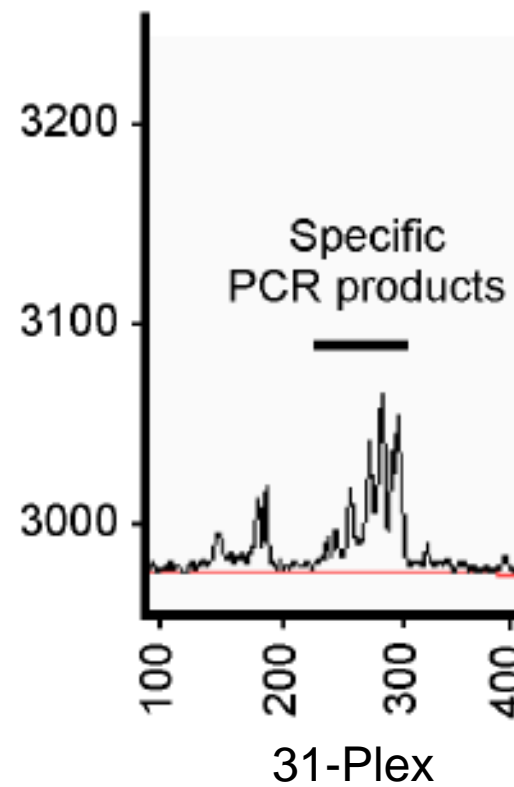
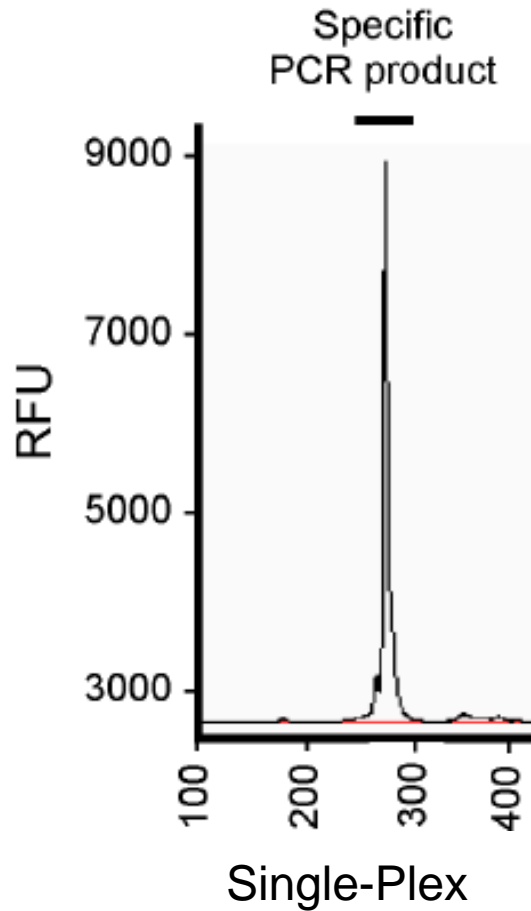
# Library Construction is Extremely Quick and Simple

- Two rounds of PCR
- Single purification step.
- Three hours from start to sequence-ready.





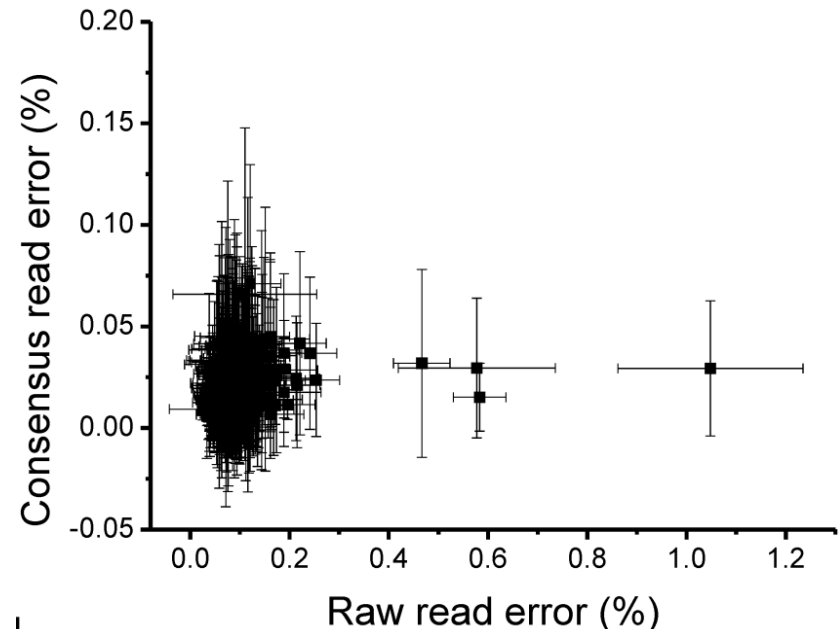
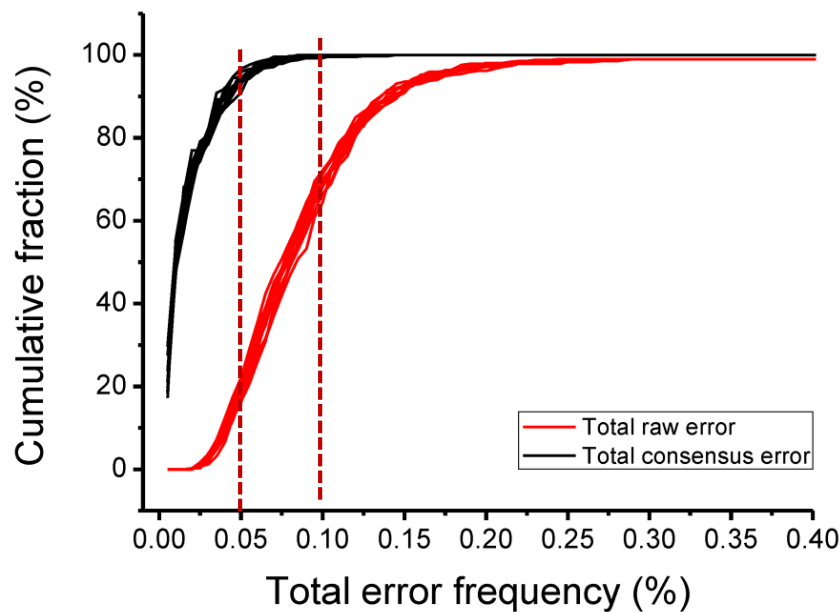
# SiMSen-Seq Enables Flexible Multiplexing



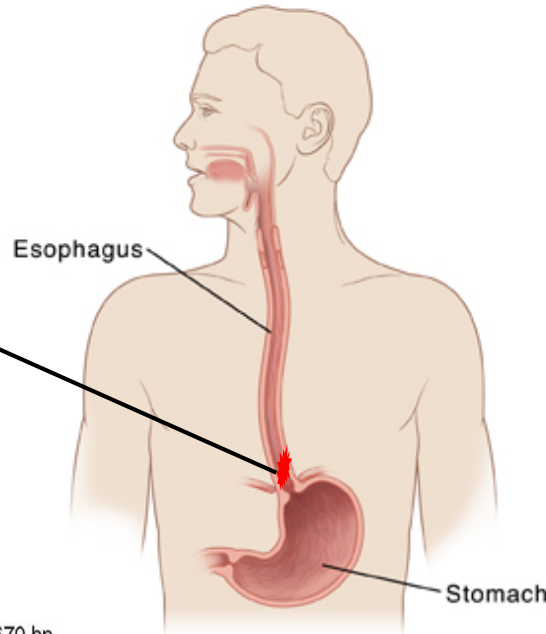
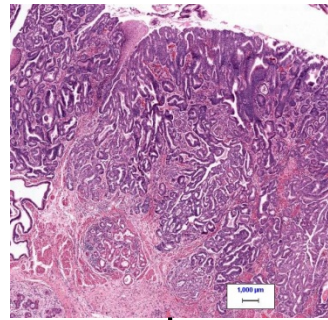
# Background Consensus Error is Consistently Below 0.1%

12 replicates of 5-plex library covering 417 nucleotides

>90% of all nucleotides displayed consensus read error <0.05% and 99.3% of nucleotides showed consensus error <0.1% with 95% confidence.

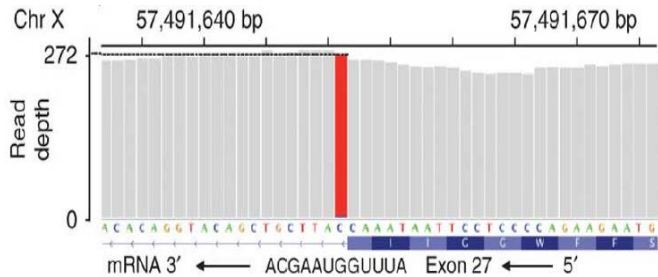


# Approach



Sequence Tumor DNA

Isolate Plasma DNA



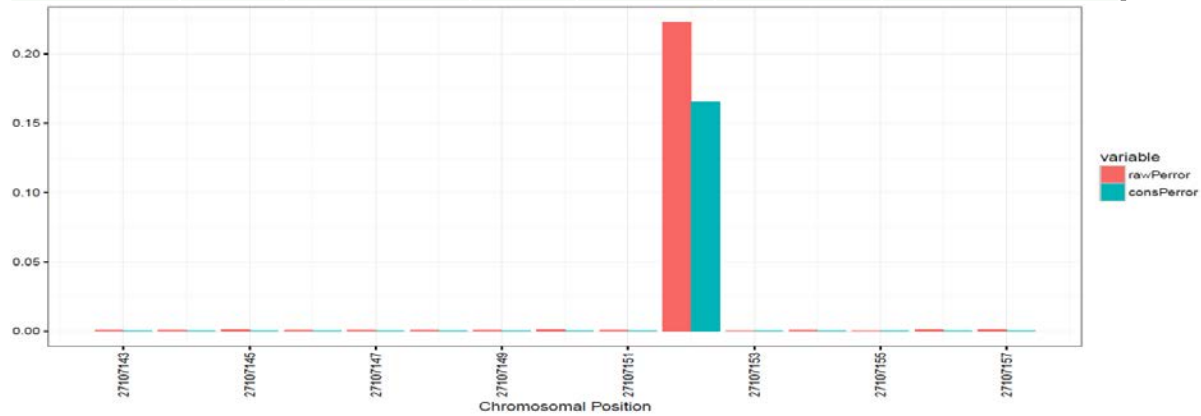
Identify Mutations

Barcoded Plasma DNA Sequencing

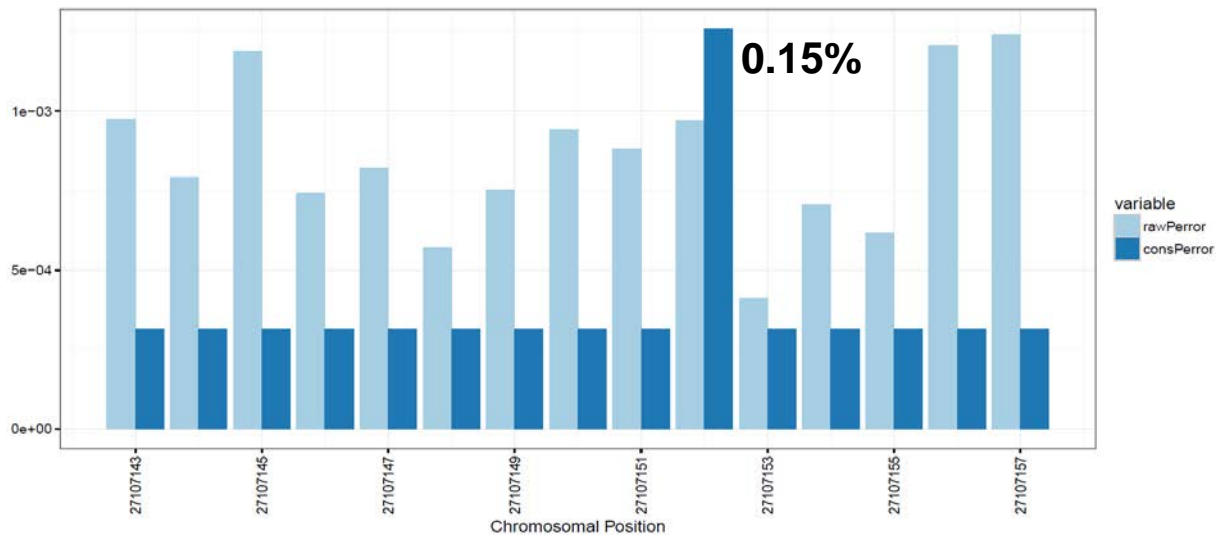
# Pitt 07: T2N0M0, Stage IB

Gene	Chr	Position	Allele 1	Allele 2	Tumor allele freq	Mutation consequence
ARID1A	Chr1	27107152	G	T	0.2	Nonsense

Tumor



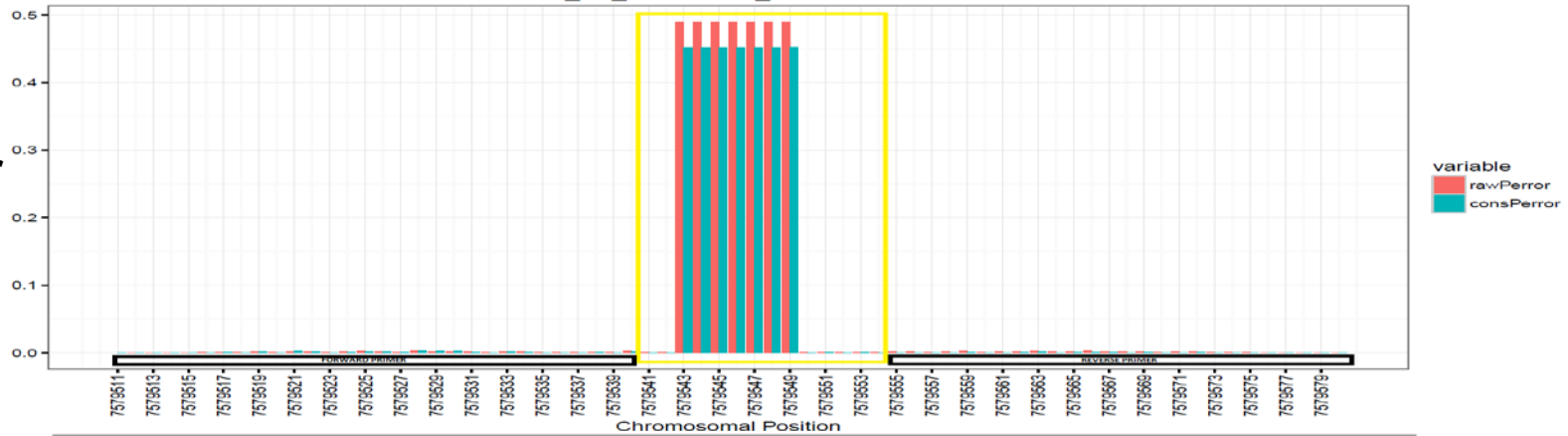
Plasma



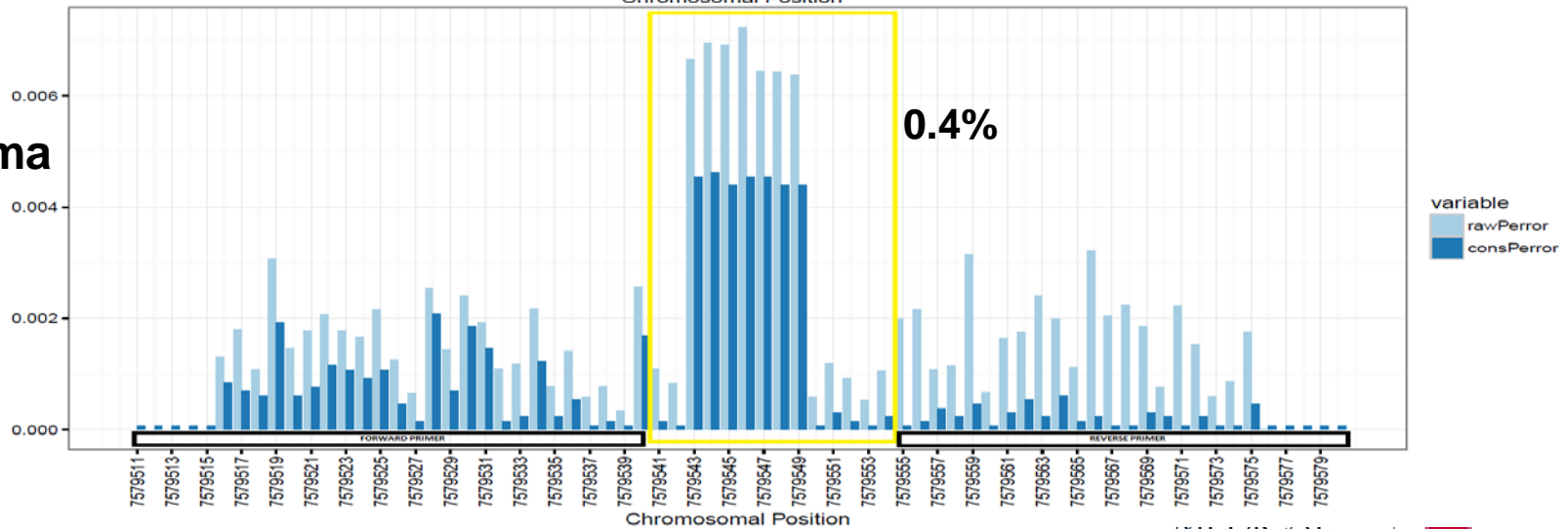
# Pitt 29: T2NOMO, Stage IIB

Gene	Start Position	Allele 1	Allele 2	Tumor allele freq	Mutation consequence
TP53	Chr17 7579542	CGTCCGGG	C	0.45	Deletion

Tumor

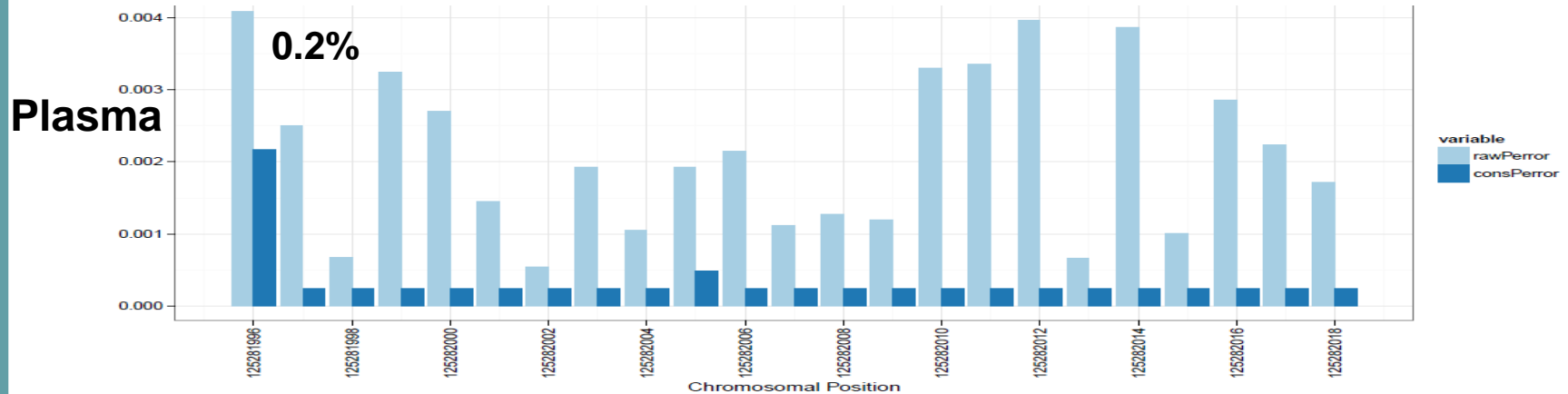
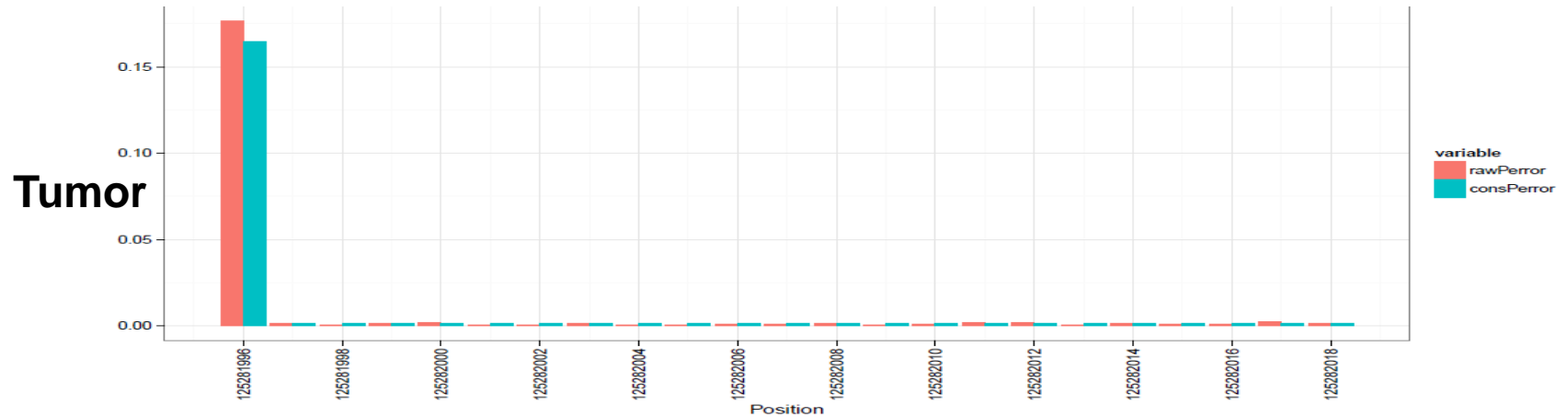


Plasma



# Pitt 27: T3N2MX, Stage IIIB

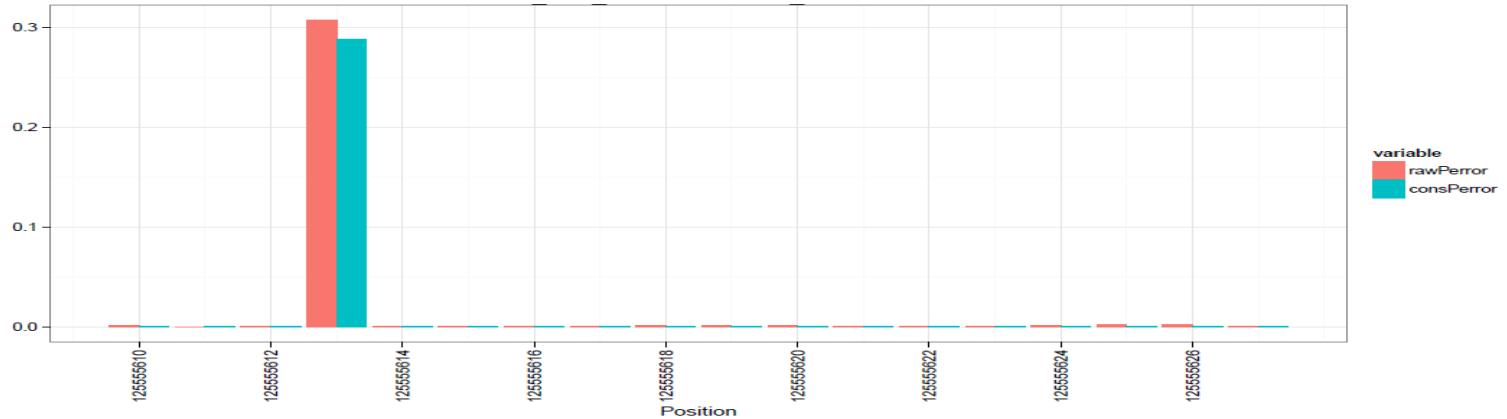
Gene	Chr	Position	Allele 1	Allele 2	Tumor allele freq	Mutation consequence
CNTNAP5	Chr2	125281996	C	A	0.16	Missense



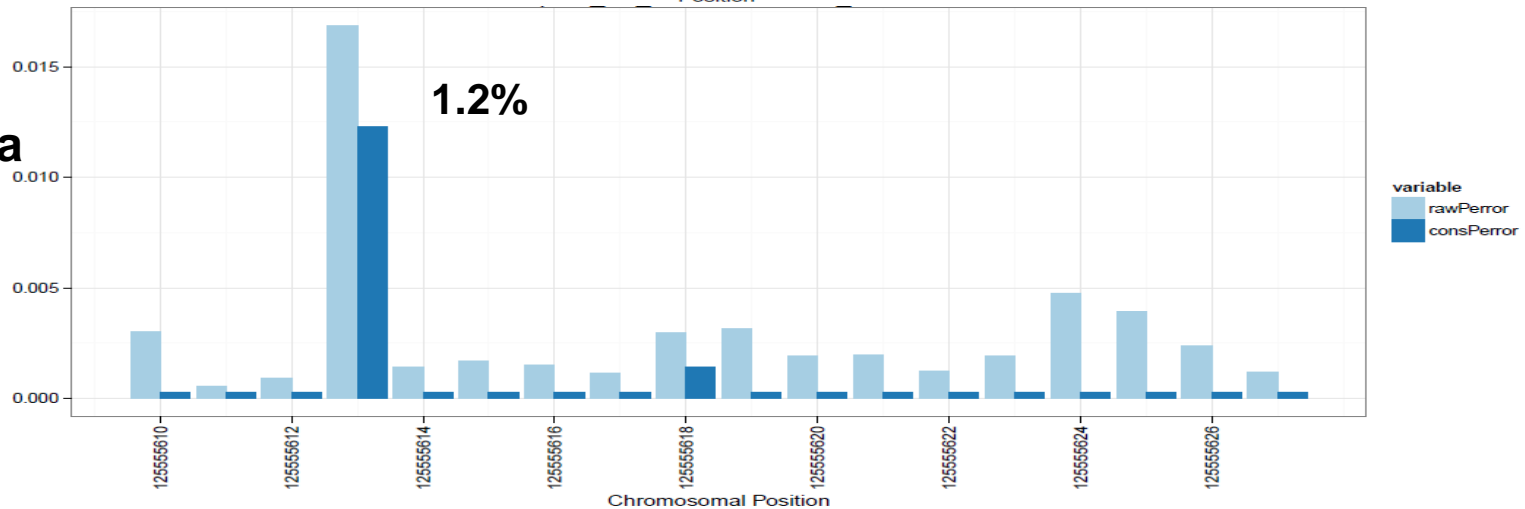
# Pitt 25: TxN2M1, Stage IV

Gene	Chr	Position	Allele 1	Allele 2	Tumor allele freq	Mutation consequence
CNTNAP5	Chr2	125281996	T	G	0.31	Intron

Tumor



Plasma

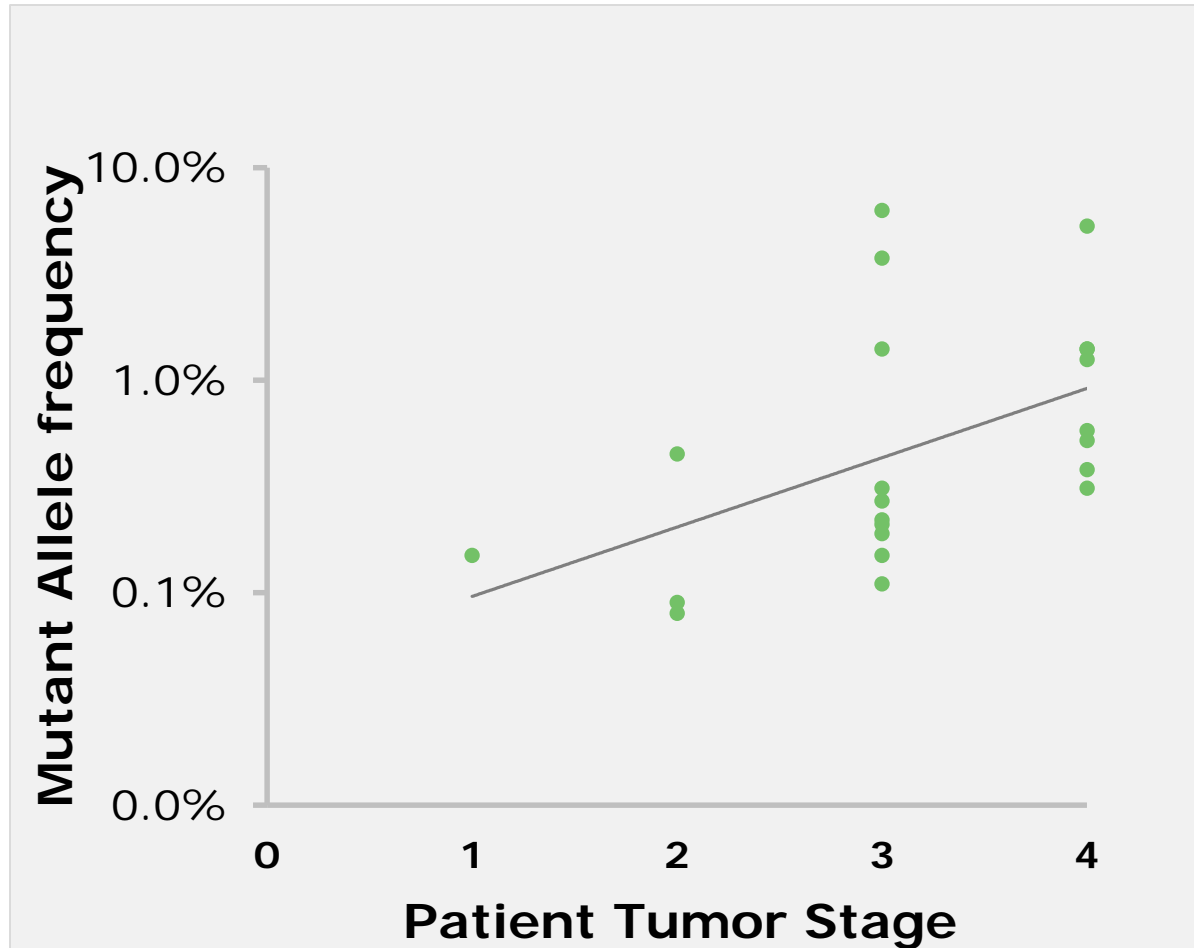


# Circulating Tumor DNA Detection Frequency Increases with Stage

Stage	Total Pts	Pts w/ tumor mutations	Plasma Sequenced	Plasma mutations detected
I	9	5	4	1 (25%)
II	16	11	5	3 (60%)
III	18	16	12	6 (50%)
IV	7	5	5	4 (80%)
<b>Total</b>	<b>50</b>	<b>37</b>	<b>26</b>	<b>14 (54%)</b>



# Mutant Allele Frequency in Plasma Increases with Tumor Stage



# Improving Detection Sensitivity

- Evolution of SiMSen-Seq:
  - Shorter amplicon sizes

# Amplicons <80bp Give Greater Sensitivity

Stage	Pts with tumor mutations	Plasma Sequenced	Plasma mutations detected	Long Amplicons Only	Short Amplicons Only
I	5	4	1 (25%)	0/1 (0%)	1/3 (33%)
II	11	5	3 (60%)	0/1 (0%)	3/4 (75%)
III	16	12	6 (50%)	3/7 (43%)	3/5 (60%)
IV	5	5	4 (80%)	0/1 (0%)	4/4 (100%)
<b>Total</b>	<b>37</b>	<b>26</b>	<b>14 (54%)</b>	<b>3/10 (30%)</b>	<b>11/16 (69%)</b>

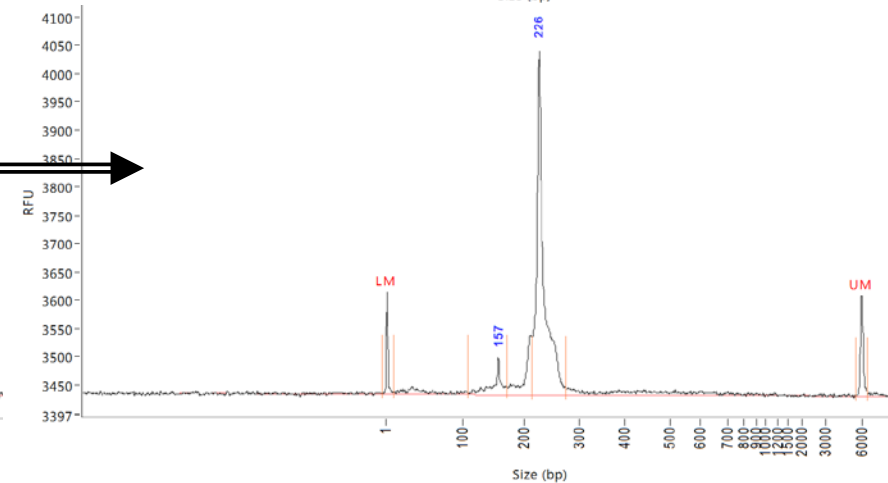
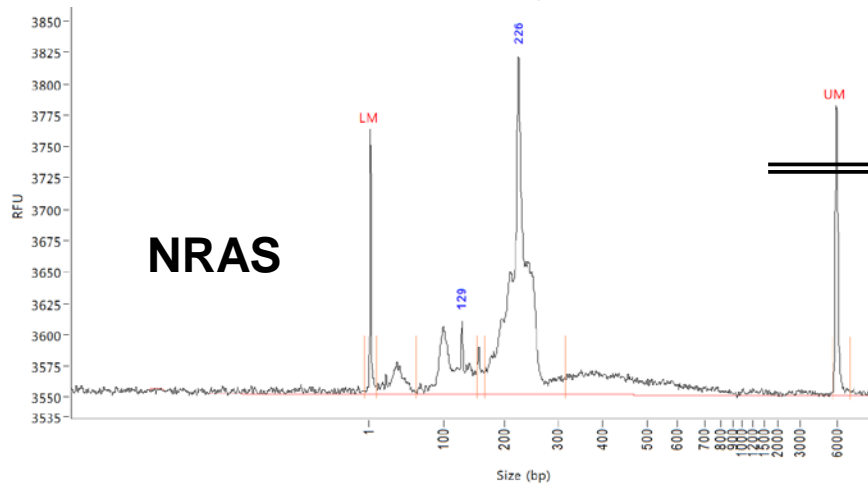
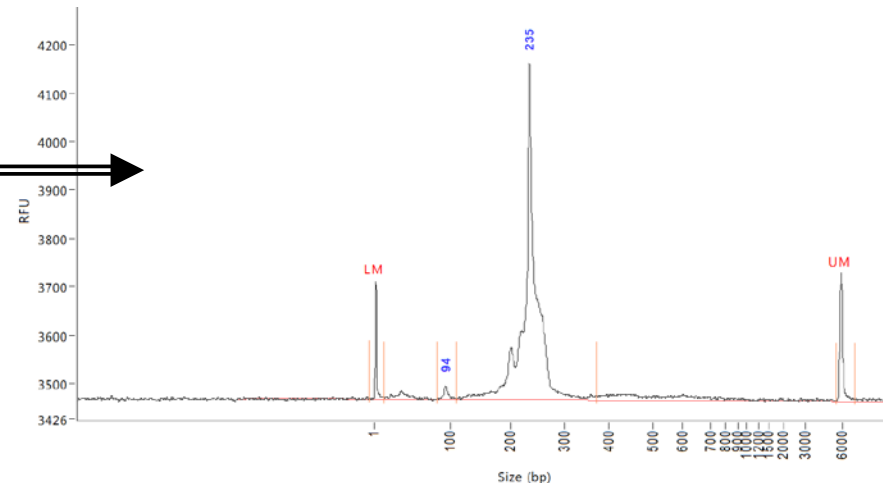
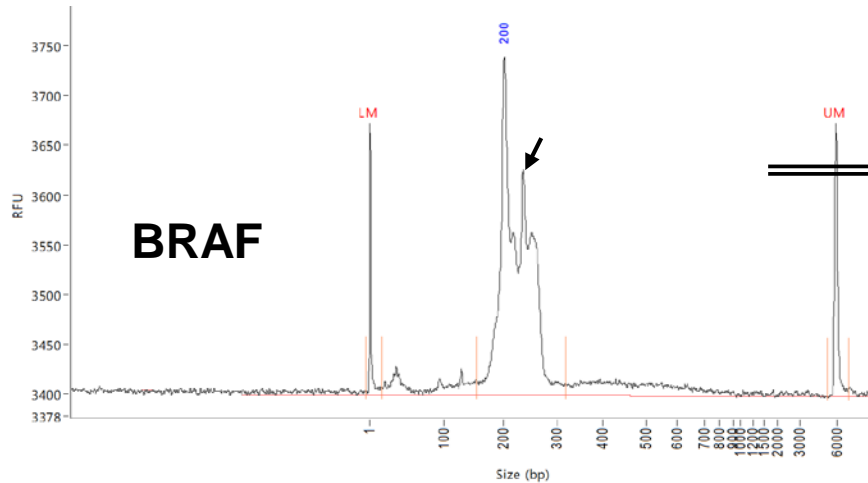
# Improving Detection Sensitivity

- Evolution of SiMSen-Seq:
  - Shorter amplicon sizes
  - PAGE purified hairpin/adaptor primers in first round PCR

# PAGE Purified Primers give Cleaner Libraries

Non-PAGE

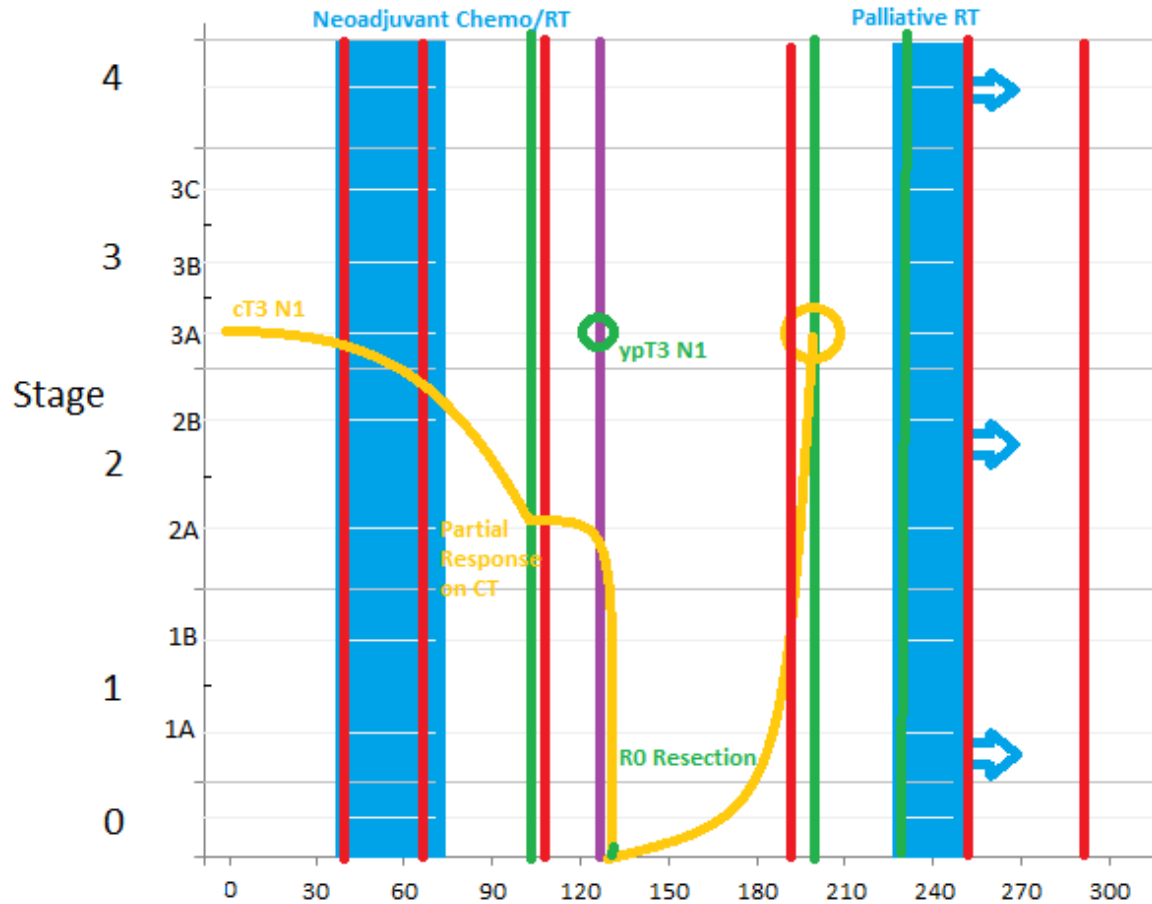
PAGE



# Improving Detection Sensitivity

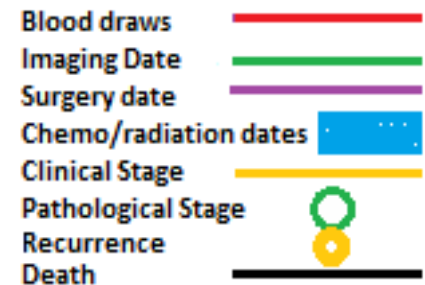
- Evolution of SiMSen-Seq:
  - Shorter amplicon sizes
  - PAGE purified hairpin/adaptor primers in first round PCR
    - Higher on-target reads and higher consensus depth
  - True Hi-fidelity polymerase in first round PCR
    - Phusion polymerase reduces background error to <0.02%

# Does ctDNA quantity change with Disease Status?

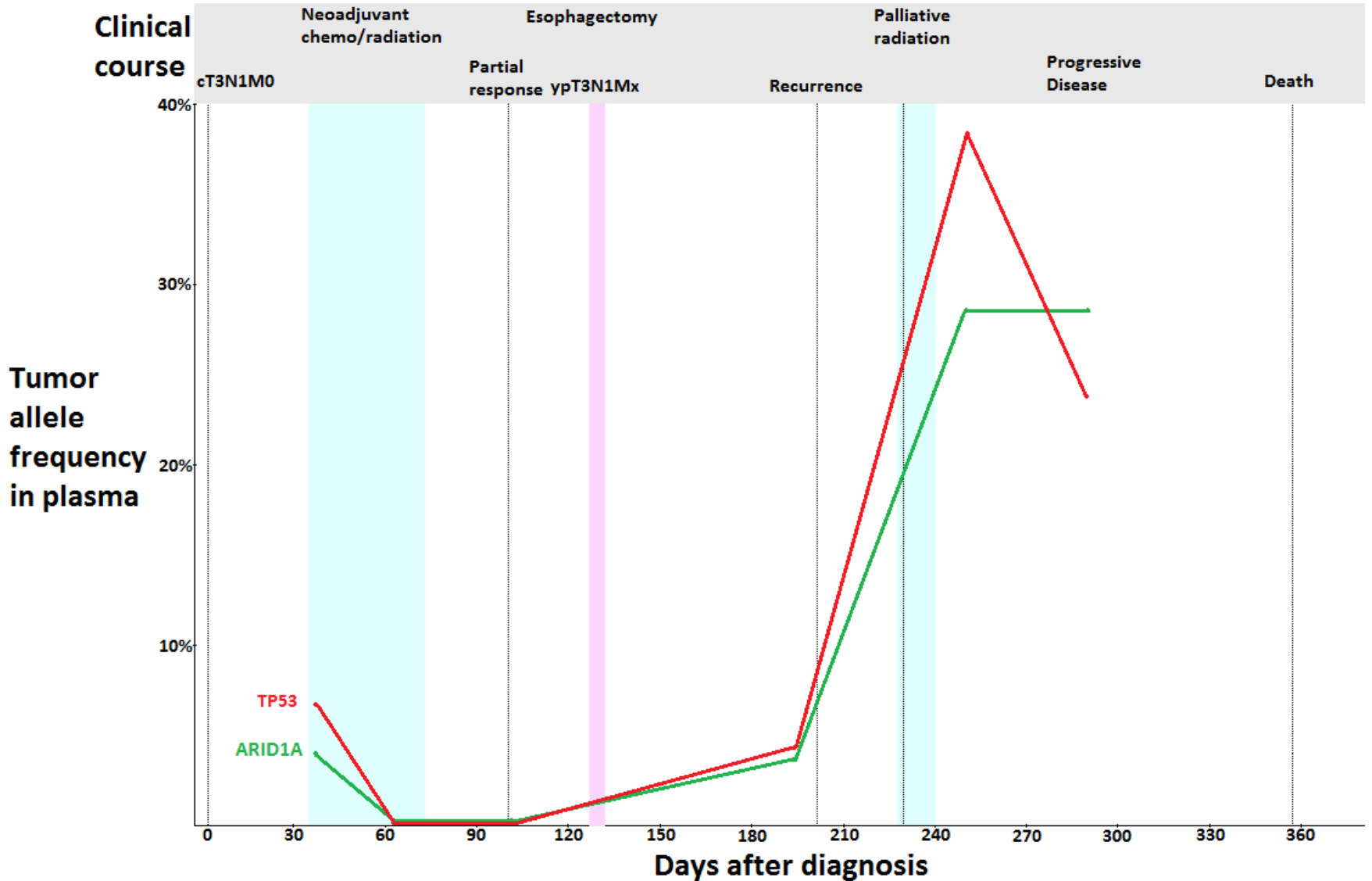


Stage IIIA patient

Point Mutations in:  
ARID1A  
TP53



# ctDNA Level Correlates with Clinical Disease Burden





# SiMSen-Seq: Strengths and Weaknesses

## •Strengths

- Easy, fast library construction and relatively simple bioinformatics
  - Implement in individual research and clinical labs
- Low DNA input requirement (<5ng)
- Flexible library content
  - Most useful for 1-1000bp coverage
  - Content can be customized easily (individual patient panels, cancer-specific panels, therapeutic panels, companion diagnostics etc.)
- Low cost for NGS approach (only sequencing regions of interest)
- Fits perfectly between dPCR and large-scale NGS approaches.

## •Weaknesses

- Up-front assay development needed
- Limited content relative to other NGS approaches
- Sensitivity not quite as good as reported for some approaches

# ctDNA as a Biomarker in Esophageal Adenocarcinoma

- Early Detection
  - Unclear if sensitivity will be high enough for stage I/II disease
    - But: High risk group known and current paradigm is failing
- Treatment response and recurrence monitoring
  - May have value for rapid identification of response to therapy
  - May identify pre-clinical recurrence
  - May identify residual disease following “curative” treatment
- Other
  - Prognostic biomarker in stage I/II disease

# Acknowledgments



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