#### Deep Sequencing of Circulating Tumor DNA for Cancer Detection and Monitoring





#### Max Diehn MD, PhD

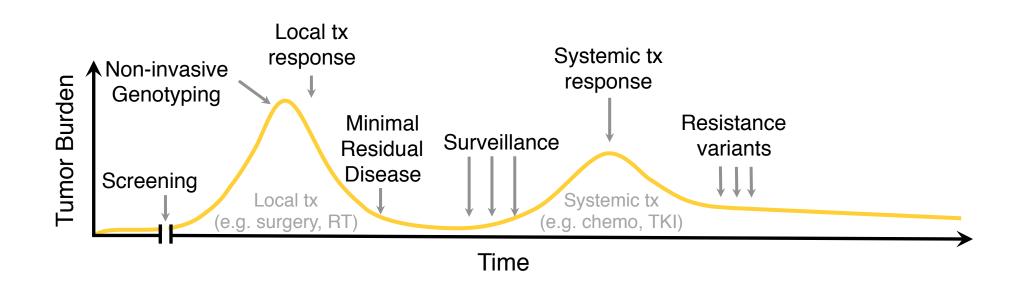
Assistant Professor of Radiation Oncology Stanford Cancer Institute Institute for Stem Cell Biology and Regenerative Medicine Stanford University School of Medicine



## **Disclosure Information**

- I have the following financial relationships to disclose:
  - Consultant for: Roche, Quanticel Pharmaceuticals, Novartis, CiberMed
  - Founder/stock holder: CiberMed
  - Grant/Research support from: Varian Medical Systems, Genzyme

### Potential Clinical Applications of ctDNA



# **Clinical Utility of ctDNA Detection**

- Clinical utility of ctDNA is largely still being established
- Utility has been documented for non-invasive genotyping
  - EGFR activating mutations in NSCLC (FASTACT-2 trial - Mok et al. CCR 2015)
- Other potential applications are in early stages of being explored
  - Monitoring treatment resistance mechanisms
  - Minimal residual disease

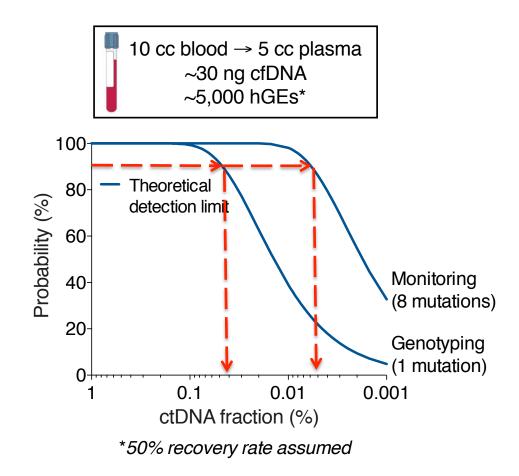
#### NGS-based ctDNA Detection

#### Comparison of ctDNA Detection Limits in 10 mL Blood Draw

ctDNA detection method	Detection limit*
Sanger sequencing	>10%
Pyrosequencing	~10%
Whole exome sequencing	~5%
Whole genome sequencing	~1%
WholeAmplicon NGS (e.g eTAm-Seq)	~0.3%
Allele-specific PCR (e.g. Intplex)	~0.1-0.05%
Barcoded Amplicon NGS (Safe-SeqS)	
Digital PCR (e.g. ddPCR, BEAMing, etc.)	~0.05-0.1%
Capture-based NGS (CAPP-Seq)	~0.00025%

\*50% efficiency, 90% probability of detection

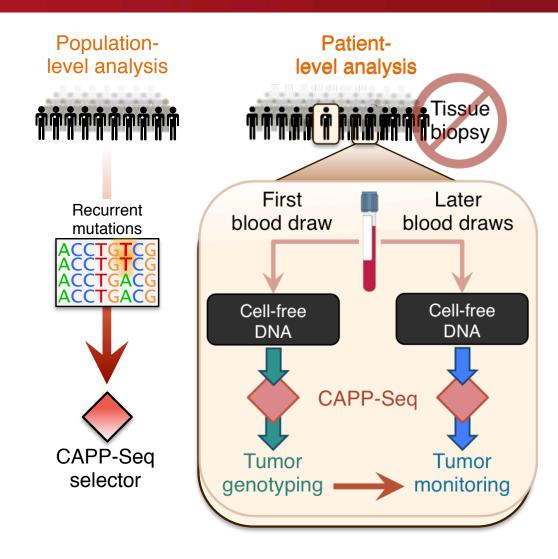
### Tracking Multiple Mutations Increases Maximizes Sensitivity



Newman, Lovejoy, Klass, et al., *Nat Biotechnol.* 2016

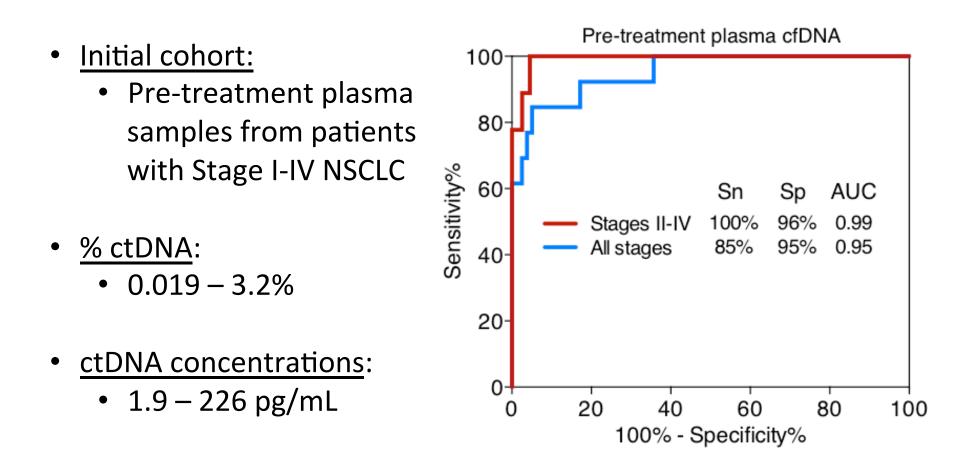
M. Diehn / Stanford

#### Cancer Personalized Profiling by Deep Sequencing (CAPP-Seq)



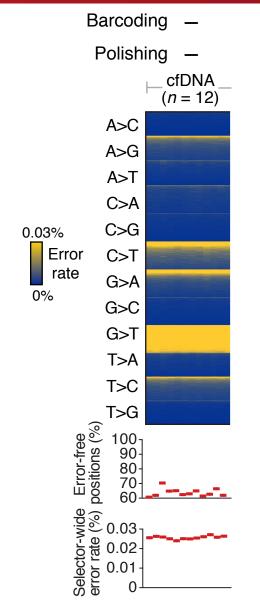
**Personalized** biomarker for every patient

#### Sensitive and Specific Detection of Circulating Tumor DNA



Newman & Bratman et al. Nature Medicine 2014

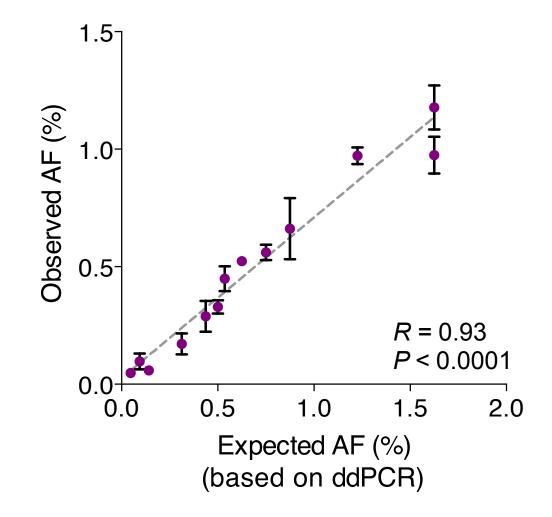
### Decreasing Sequencing Errors in Deep Sequencing-based cfDNA Analyses



= <u>integrated</u> <u>Digital</u> <u>Error</u> <u>Suppression</u>

Newman, Lovejoy, Klass, et al., *Nat Biotechnol.* 2016

#### Comparison of iDES-enhanced CAPP-Seq to Digital PCR



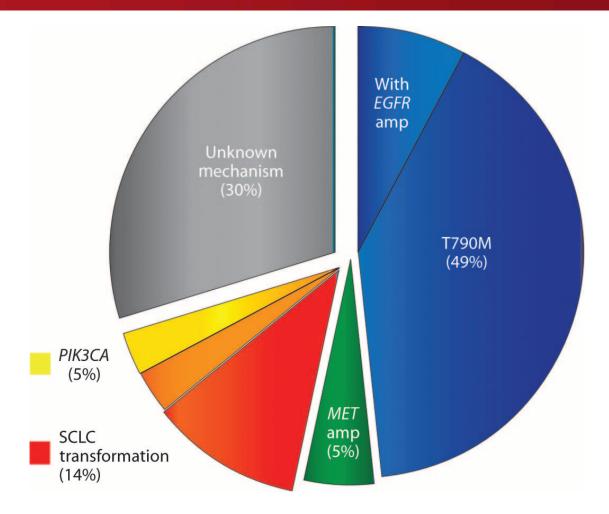
#### Resistance Mechanisms in EGFR Mutant NSCLC

# **Targeting EGFR Mutations in NSCLC**

- Activating EGFR mutations occur in 15-50% of lung adenocarcinomas
- Sensitize tumors to EGFR tyrosine kinase inhibitors
  - First line: erlotinib, gefitinib, afatinib
- Resistance invariably develops

   EGFR T790M is most frequent mechanism (~50-60%)
- "Third generation" EGFR TKIs target both activating and T790M mutations
  - Osimertinib, rociletinib, and others

#### Frequency of First-line EGFR TKI Resistance Mutations in Tumor Biopsies

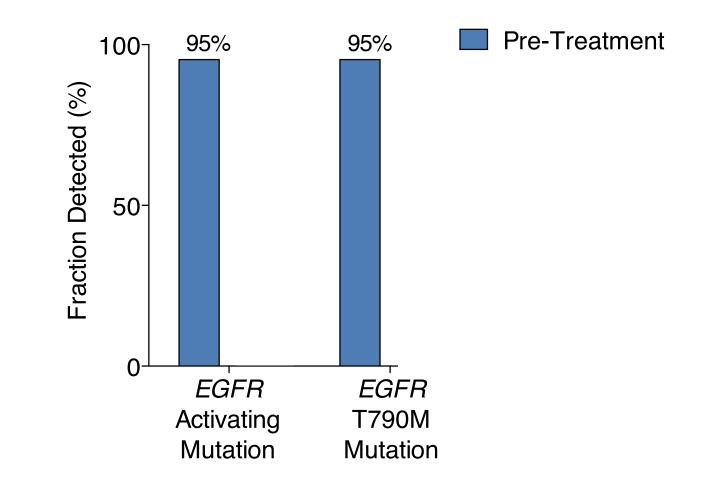


~5-15% of patients with more than one mechanism

# Heterogeneity of Resistance Mechanisms in Response to EGFR TKIs

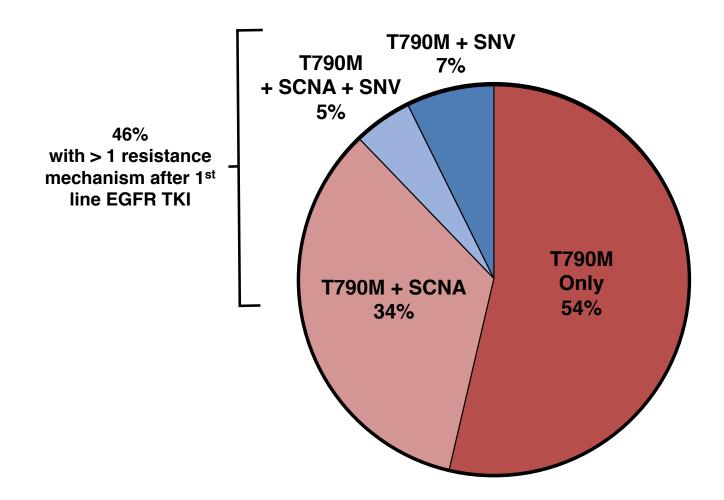
- Hypotheses
  - First-line EGFR TKI treatment frequently leads to <u>intra</u>patient heterogeneity in resistance mechanisms
  - Patients with multiple resistance mechanisms respond less well to third generation EGFR TKIs
- Approach
  - Perform CAPP-Seq on plasma from 43 patients who progressed on first-line EGFR TKIs and were subsequently treated with rociletinib
  - Analyze baseline and emergent resistance-associated somatic alterations

#### ctDNA Detection Summary



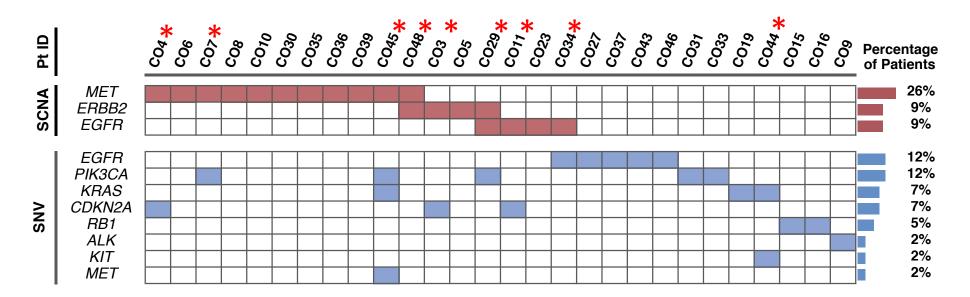
Chabon et al. Nature Communications 2016

# Intra-patient heterogeneity of resistance mechanisms to first-line EGFR TKIs



#### **Rociletinib Resistance Mechanisms**

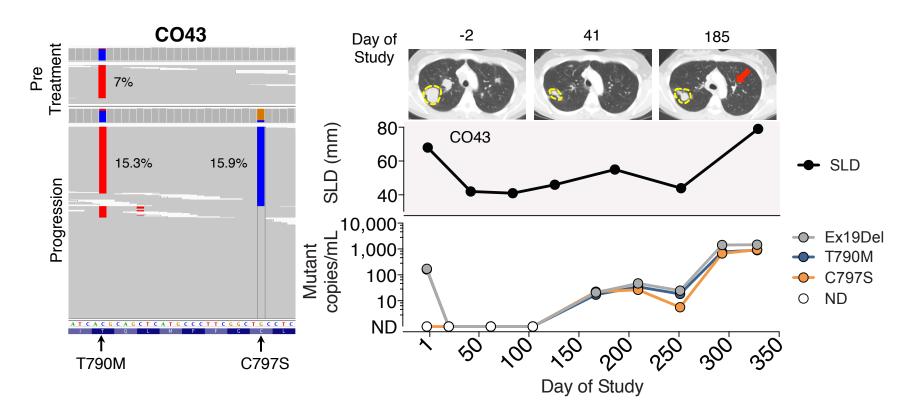
- Putative resistance mechanism definition
  - Absent before treatment and emerged at progression
  - Increased in relative abundance from baseline to progression
- Putative mechanism(s) identified in 65% of patients (72% of evaluable)
- Significant intra- and inter-patient heterogeneity
  - 9 genes involved
  - 21% of patients develop multiple resistance mechanisms (\*)



#### Emergence of EGFR C797S in

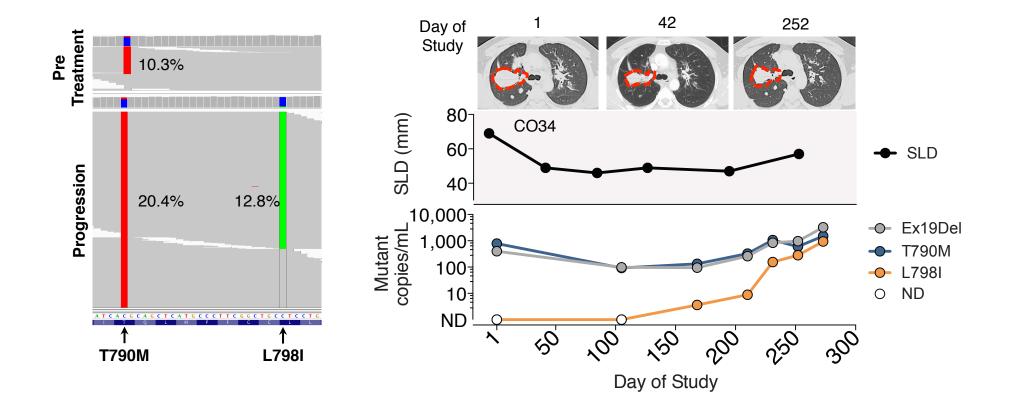
 Inress et al.
 Plasma
 osimertinib
 19
 32%

 a simertinib
 Taucini

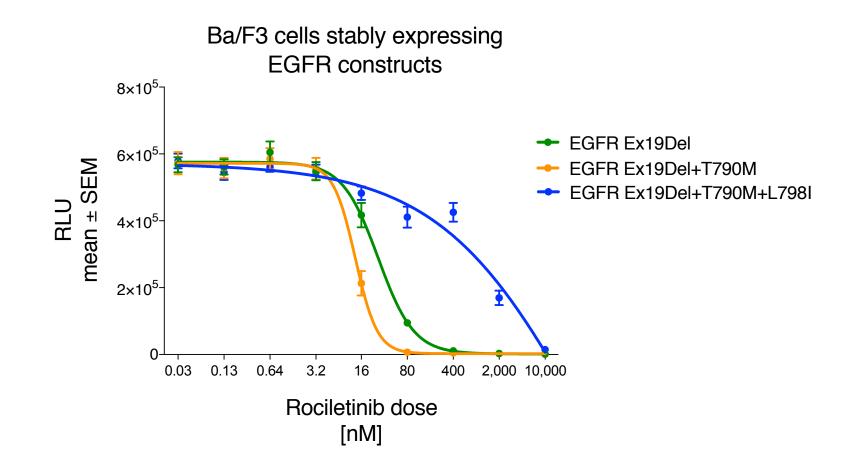


Study	Compound	Evaluable Patients	C797S +	C797S Prevalence
Present Study Piotrowska <i>et al.</i> Cancer Discovery 2015	Rociletinib	40 9*	1 0	2%

#### Novel EGFR L798I Resistance Mutation

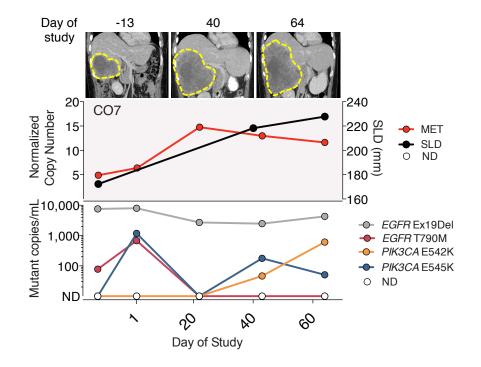


#### EGFR L798I Mutation Causes Rociletinib Resistance

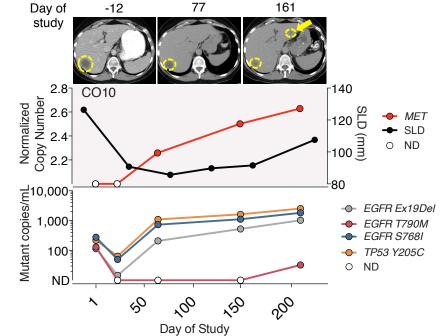


#### MET amplification mediates innate & acquired resistance

#### **Innate Resistance**



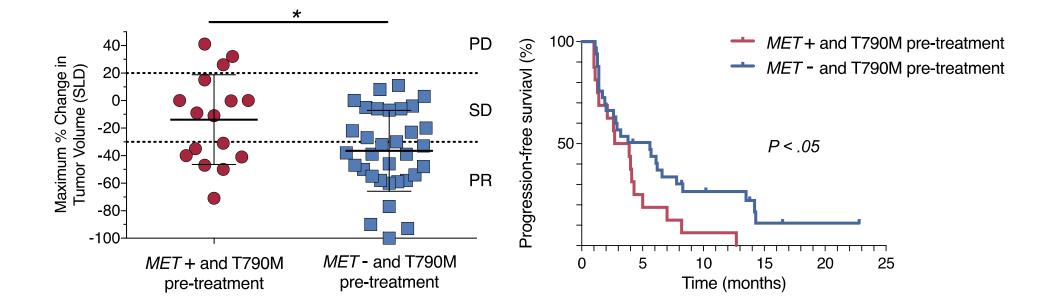
#### **Acquired Resistance**



### Presence of Multiple Resistance Mechanisms predicts poor outcome

#### **Expanded MET Cohort**:

- 16 patients with T790M+/MET+
- 33 patients with T790M+/MET-



# Summary

- ctDNA analysis has many potential clinical applications
- NGS-based methods such as CAPP-Seq can achieve similar sensitivity as ddPCR and facilitate broad molecular profiling and monitoring
- Simultaneous monitoring of multiple resistance mechanisms may allow personalized targeting of emerging resistance mutations
- Detection of MRD requires ultrasensitive assays and may allow personalized therapy
  - Randomized trial in Stage II colorectal cancer ongoing (J. Tie et al.)
- More prospective clinical studies required to validate preliminary findings and to establish clinical utility

### Acknowledgements

CAPP-Seq Team				
PIs: <u>Max Diehn &amp; Ash Alizadeh</u>				
Aaron Newman	Florian Scherer			
Alex Lovejoy	Leslie Modlin			
Dan Klass	Evan Osmundso			
Henning Stehr	David Kurtz			
Jake Chabon	Chih Long Liu			
Angela Hui	Carmen Say			
Aadel Chadhuri	Justin Carter			
Li Zhou	Alexander Craig			

Youngtae Jeong Horace Rhee Luke Lee Angela Hui Sharavani Sinha Jeremy Harris Andrew Gentles



#### <u>Funding</u>

n

NIH New Innovator Award NIH-NCI: R01, U01 DOD **Other collaborators** Bill Loo David Shultz Michael Gensheimer Heather Wakelee Joel Neal Joseph Shrager Mark Berry **Robert Merritt Robert West** Carmen Say **Justin Carter** Andy Simmons **Chris Karlovich** 

Doris Duke Foundation V Foundation CRK Research Fund